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# Molecular and structural insights of $\beta$ -boswellic acid and glycyrrhizic acid as potent SARS-CoV-2 Envelope protein inhibitors

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#### ABSTRACT

Background: Over million people have been infected with SARS-CoV-2 virus worldwide, with around 3% reported deaths till date. A few conventional antiviral treatments have been tried to mitigate the coronavirus. However, many alternative therapeutics are being evaluated worldwide. In the present study, we investigated traditional Indian medicinal compounds antiviral potencies as an effective drug for targeting SARS-CoV-2E. SARS-CoV-2 E protein plays a key role in coronavirus life cycle and is an interesting target for the development of anti-SARS-CoV-2 E drugs.

*Methods*: Molecular docking studies of medicinal compounds possessing wide range of pharmacological and antiviral activities against enveloped viruses were evaluated with the computer-aided drug design screening software; PyRx. Twelve medicinal compounds isolated from plants were screened and visualized on Biovia Discovery-Studio. Moreover, SARS-CoV-2 E protein's secondary structural insights were deciphered using Swiss Model and ProFunc web server.

Results: Glycyrrhizic acid, triterpene glycoside isolated from plants of Glycyrrhiza (licorice) showed interactions with envelope protein at chain A: Arg 61, chain B: Phe 23, chain B: Tyr 57, and chain C: Val 25.  $\beta$ - boswellic acid, an ayurvedic herb (pentacyclic terpenoid are produced by Boswellia) represented direct interactions and indirect binding with chain C. Their pharmacological aspects and drug-likeness properties were deduced by DruLiTo. Toxicological assessment, along with their ADME profiling, was validated using vNNADMET. The findings showed that ligands,  $\beta$ -boswellic acid, and glycyrrhizic acid possessed the best bindings, with the target having binding affinity (-9.1 kcal/mol) amongst compounds tested against SARS-CoV-2 E. In-vitro studies reveals the promising effect as potent SARS-CoV-2 E inhibitors. Functionality loss and structural disruptions with  $\sim$ 90% were observed by UV-spectra and fluorescent based analyses.

Conclusion: The study demonstrated that  $\beta$ -boswellic acid, and glycyrrhizic acid are strong SARS-CoV-2 E protein inhibitors. In addition, the work linked GA antiviral activity to its effect on SARS-CoV-2 E protein that can pave the way for designing antiviral therapeutics.

# Introduction

The disease produced by SARS-CoV-2 virus continued to rise, and on March 11, 2020, the World Health Organization declared COVID-19 as a pandemic (WHO, 2020). The novel Coronavirus is of zoonotic origin, mutated, and then inflicted humans with devastating outcomes (Collins, 2020, Mousavizadeh and Ghasemi, 2021). The structure of SARS-CoV-2 consists of spike (S), membrane (M), envelope (E), and nucleocapsid (N) four main structural proteins (A. Wu and Chiwaya, 2020). Moreover, there are three to four viral proteins in the coronavirus membrane

(Mousavizadeh and Ghasemi, 2021). The SARS-CoV-2 E protein is a minor but essential component of the SARS-CoV-2 virus. It is a small integral membrane, pentameric (A-E chains) that forms the ion channel viroporin (Alam et al., 2020; Mandala et al., 2020; Sarkar and Saha, 2020). It is composed of 75–109 amino acid integral membrane protein, hydrophilic N-terminal domain (7–12 residues). It also has a long hydrophilic C-terminal domain, is mostly  $\alpha$ -helical. It features a single hydrophobic transmembrane domain (HD) made up of 25 residues, targeted to Golgi membranes for virion release and secretory pathway cascade (Day, 2020; Surya et al., 2018).

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While S protein was the focus of much research efforts, recently, other proteins including E proteins are also thoroughly investigated (Bojadzic et al., 2020; Caliebe et al., 2021; Li et al., 2021; Yu et al., 2021). The work by Caliebe et al. (2021) reported bioinformatics analysis of three major Boswellia resin constituents and discovered that they bind to functional proteins viz. the replicase polyprotein PODTD1, the spike glycoprotein PODTC2, and the nucleoprotein PODTC9 of the SARS-CoV-2 virus. Moreover, Li et al. (2021) demonstrated by computational simulation data that glycyrrhizic acid interacted with S protein for its antiviral activity, thereby blocking SARS-CoV-2 infection. Moreover, the significance of SARS-CoV-2 E protein is to maintain a conserved Golgi complex-targeting signal being essential for the development of virus life cycle, (Day, 2020) including the processes of assembly, budding, envelope formation, and pathogenesis (Alam et al., 2020; Collins, 2020; Sarkar and Saha, 2020). It has been discovered to be a virulence factor and to play a role in virus pathogenesis (Javorsky et al., 2021). The cationic activity viroporins of CoV-2 E protein are the key driving factor that substantially impacts the secretory pathway governing viral productivity at the endoplasmic reticulum-Golgi intermediate compartment (ERGIC). (Collins, 2020; Day, 2020). The E protein is in a region where interactions with other CoV proteins and host cell proteins occur (Sarkar and Saha, 2020). Hence, it stands as a crucial link in COVID-19 transmission (Roy and Menon, 2021).

On the basis of studies gained from SARS and MERS infections, reports on the use of antiviral drugs, hypertensive medicines, or even anti-inflammatory drugs or angiotensin-converting enzyme (ACE) inhibitors along with angiotensin-receptor blockers (ARBs). Based on research on SARS, MERS, and Ebola outbreaks, scientists predict that conventional antiviral drugs like Remdesivir (nucleoside analogs), Carprofen, and Celecoxib (anti-inflammatory drugs) or Lopinavir/Ritonavir (anti-retrovirals) could benefit from curing COVID-19 as well.

The urgency is to come up with an alternative solution to control and break the chain of COVID-19 transmission. Traditional Indian medicinal plants have shown to be promising source of compounds to cope COVID-19 disease (Alam et al., 2020; Saravanan et al., 2020). To discover the potent plant metabolites/molecules for the treatment and prevention of COVID-19, traditional plant secondary metabolites' screening is highly recommended. Our previous work based on screening the novel medicinal metabolites that could be a target for SARS-CoV-2 E protein (Alam et al., 2020).

The novelty of the present study was to find compounds that are components of some Indian medicinal plants that could inhibit SARS-CoV-2 E protein by molecular docking, along with in-vitro assays.

Hence, this study proves to provide a promising approach for preventing COVID-19 that remains to be deciphered further. Understanding the role of the SARS-CoV-2 E protein in viruses is thus a paramount prerequisite for identifying novel antiviral therapeutics.

### Materials and methods

Chemicals

SARS-CoV-2 Envelope protein, His-and Avi tag (Cat No. GTX01547-pro) was purchased from GeneTex, Inc., CA, USA. All other reagents used were of analytical grade.

Selection of compounds that are present in Indian medicinal plants

Twelve compounds from different Indian medicinal plants (Table 1) were selected from a library of Indian plants named the IMPPAT database, and the compounds and their structures were taken from PubChem (http://pubchem.ncbi.nlm.nih.gov). All the structures were converted from Smiles strings to PDB file format using Chimera software tool 12.1 (Pettersen et al., 2004).

Structural insights of the target SARS-CoV-2 E protein

SARS-CoV-2 E protein was used as a template (PDB ID-7K3G) (Mandala et al., 2020). Ramachandran plot was analyzed and showed the energetically favoured regions for backbone dihedral angles against amino acid residues in protein structure. ProFunc webserver was used to decipher the target protein's overall structure and function (htt p://www.ebi.ac.uk/thornton-srv/databases/ProFunc).

Docking studies of SARS-CoV-2 E protein

Molecular docking was performed on SARS-CoV-2 E of SARS-CoV-2 imported from Protein Data Bank (https://www.rcsb.org/), as PDB ID-7K3G (Mandala et al., 2020)". The ligand structures were obtained from PubChem.Significant pockets and groves present in the target proteins were visualized using Caver Analyst 2.0 beta. Docking studies of target molecules with envelope proteins of SARS-CoV-2 were performed using the PyRX virtual screening tool (Dallakyan and Olson, 2015; Trott and Olson, 2020). The input file format was a PDB file. The binding affinity of the ligand with target proteins was studied, and compounds with better binding affinity were determined. Docking results and interaction of critical amino acid residues were shown and visualized using Biovia Discovery Studio

Pharmacological profiling of medicinal compounds

The drug likeliness of the twelve selected compounds was checked with DruLiTo1 (Westerbeck and Machamer, 2019). ADMET prediction, toxicity, and recommended dose of screened herbal compounds were estimated with the vNN ADMET prediction tool.

Inhibition of SARS-CoV-2 by the twelve selected natural compounds from indian medicinal plants monitored with UV-visible spectra

The inhibitory effects of the twelve selected natural compounds on the activity of SARS-CoV-2 were tested. To monitor the structural and functional changes, UV–vis scans in the range 200–700 nm were obtained with a Shimadzu UV-1800 spectrophotometer double beam. The absorbance values were obtained by keeping the SARS-CoV-2 E protein concentration constant while varying concentrations of  $\beta$ -boswellic and glycyrrhizic acids (from 50 to 100 µg/ml at pH=7.4) were successively added.

Fluorescence spectroscopic measurements

The fluorescence emission spectra were acquired on Perkin Elmer LS50 fluorospectrometer (Perkin Elmer, Boston, MA, USA) equipped with  $1.0~\rm cm$  path length quartz cells at  $25~\rm ^{\circ}C$ . The emission spectra were recorded between 310 and 450 nm after excitation at 295 nm and slit widths were kept at 7 nm and 10 nm for excitation and emission respectively. The spectra were recorded in triplicate and normalized for buffer contributions.

Statistical analysis

All the experiments were performed in triplicate, and data were expressed as mean  $\pm$  standard deviation (SD) from replicate runs. Analysis of variance (ANOVA) using Origin 2020 (9.7) (OriginLab Corporation, Inc., Northampton, MA, USA) was considered to indicate a statistically significant differences between samples (p < 0.05).

### Results and discussion

Selection of compounds from Indian medicinal plants for SARS-CoV-2 E inhibitors

In our study, the twelve compounds detailed in Table 1 were selected from traditional Indian medicinal plants. These compounds were

screened based on their reported antiviral and anti-inflammatory properties (Benarba and Pandiella, 2020) and thus, they can be promising antiviral drugs against SARS-CoV-2. These compounds are used in the treatment of various respiratory diseases, which are the primary targets in COVID-19 infections (Ahmad et al., 2021; Patel et al., 2021; Verma et al., 2021). The conventionally used commercial antiviral drugs in the clinics were also considered relevant to the present study (Table 1) (Bojadzic et al., 2020).

Structure-based pharmacophore modeling

The detailed structure elucidation of the target E protein, assessing

backbone was studied. Histogram of Ramachandran plot used to count  $\Phi$  (Phi; C—N-CA-C) /  $\Psi$  (Psi; N—CA-C-N) occurrences displayed categories A to D in Fig. 1.

The plot distribution favored 88.93% amino acid with 16.8% rotamer outliers and 2.14% Ramachandran outliers. ProFunc webserver was used to examine the overall structure and functions of the E protein. Secondary structure prediction of single-chain revealed  $\alpha$ -helix (46.6%),  $3_{10}$  helix (13.8%) and others (39.7%). It also depicted secondary structural elements like fifteen helix-helix interaction, six helices, seven  $\beta$  turns and two  $\gamma$  turns. Major secondary structures are highlighted in Fig. 2.

Further to it, the interactions among subunits are deciphered. The

**Table 1**Natural compounds from Indian medicinal plants and standard antiviral drugs that have been screened against SARS-CoV-2 E protein.

Name of the compound	Plant Source	Chemical Structure	Molecular Weight	Binding Affinity (- kcal/mo l)	Root Square Mean Deviation (RMSD)
Aloesin	Aloe vera (L.) Burm. f (Xanthorrhoeaceae)		394.13	6.4	14.53
Azadirachtin	Azadirachta indica (L.) A. Juss (Meliaceae)	OH OH	720.26	6.8	1.91
β- boswellic acid			<b>456.36</b>	9.1	1.47
Cholecalciferol	-	H,C CH <sub>0</sub>	384.34	8.8	1.6
Cinnamaldehyde	Cinnamomum verum (L.) J. Presl (Lauraceae)		132.06	6.2	12.18
Diosgenin	Trigonella foenum -graecum Linn. (Fabaceae)	CH <sub>5</sub> CCH <sub>5</sub>	414.31	8.5	1.65

(continued on next page)

Table 1 (continued)

Germacranolide	Artemisia pallens Wall. ex DC. (Compositae)	CH <sub>3</sub>	266.1	6.8	3.14
		OH <sub>3</sub>			
Gingerols	Zingiber officinale (L.) Roscoe (Zingiberaceae)	H,C OH O CH <sub>3</sub>	294.18	7.7	1.60
Glycyrrhizic acid	Glycyrrhiza glabra (L.) (Poacaeae)	H <sub>2</sub> C CH CH H <sub>2</sub> C CH H <sub>3</sub> C CH	822.4	9.1	1.85
Isofraxidin	Acanthopanax senticosus (Rupr. & Maxim.) Harms (Araliaceae)	H <sub>3</sub> C CH <sub>3</sub>	222.05	5.4	2.57
Thymoquinone	Nigella sativa (L.) (Ranunculaceae)	CH <sub>3</sub>	164.08	5.3	2.9
α-Tocopherol	-	HC————————————————————————————————————	416.37	7.7	1.64
		B. Non-natural standard drugs			

(continued on next page)

Table 1 (continued)

Azithromycin	-	H <sub>1</sub> C CH <sub>5</sub> H <sub>2</sub> C CH <sub>5</sub> H <sub>3</sub> C CH <sub>5</sub> H <sub>4</sub> C CH <sub>5</sub> H <sub>4</sub> C CH <sub>5</sub> H <sub>4</sub> C CH <sub>5</sub>	748.51	5.9	1.82
Dexamethasone	-	Ho HoH <sub>3</sub> C OH	392.2	7.2	18.56
Hydroxychloroqui ne	-	CH <sub>5</sub> CH <sub>5</sub>	335.18	6.4	4.53
Remdesivir	-	NH <sub>2</sub> N HO OH	602.23	7.7	2.0

critical amino acid residues showing interaction among chains were shown in Fig. 3.

Such interactions were found relevant during ligand interaction with the E protein. To gain insights into the structure of the SARS-CoV-2 E protein, the cavity/pocket required for catalytic sites was studied using Caver Analyst 2.0 beta. The SARS-CoV-2 E protein had cavities/pockets that were found to be essential for catalysis. A ligand-binding site was observed on the site present on the cavity. Out of twelve screened

compounds, we have demonstrated the best bindings were showed by  $\beta\text{-boswellic}$  and glycyrrhicic acids

Cavities and pockets of the proteins, as demonstrated, was targeted by these two ligands. Furthermore, structural guided studies were carried out to predict the orthosteric or allosteric sites present in the target molecules. As *per* our findings, the binding had taken place at the allosteric site. Binding at the allosteric site provides information about the nature of ligand binding. For the allosteric binding, ligands will induce

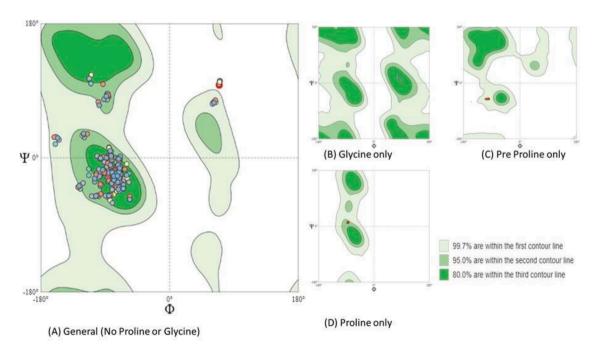


Fig. 1. Histograms of Ramachandran plot showing  $\Phi$  (Phi; C—N-CA-C) /  $\Psi$  (Psi; N—CA-C-N).

conformational changes in the protein functionality and overall protein structure (J. Wu and Chiwaya, 2020). The study also revealed that various critical residues were involved in interacting with the ligand. Various interactions such as van der waals, hydrophobic, alkyl, hydrogen bonding was visualised using Biovia Discovery Studio 2020. (Fig. 4A and B).

#### Molecular docking-based virtual screening

To further examine and visualize the binding mode of SARS-CoV-2 E protein with the screened compounds, molecular docking was performed in E protein to analyze the binding details. We analyzed the ligand's docking with the selected molecules i.e., E protein using Autodock Vina to obtain a better overview of binding affinity when the outcome was established along with root square mean deviation values (Trott and Olson, 2010). The binding affinity should be higher than 6.0. Contrary to expressing high values by binding affinity, root mean square deviation (RMSD) values need to be lowest, proving the best fit result. Table 1 shows the binding affinity of the selected plant compounds. The docking and binding affinity of the compounds with the SARS-CoV-2 E protein.  $\beta$ -boswellic acid present in the Indian plant Boswellia serrata Roxb. ex Colebr. (Burseraceae) (Fig. 4A) and glycyrrhizic acid present in Glycyrrhiza glabra L. (Poacaeae) (Fig. 4B) were found to be the best antiviral compound among the screened ligands for drug targeting of the E protein. These two unconventional metabolites of medicinal compounds showed intermolecular interaction with the SARS-CoV-2 E protein. Glycyrrhizic acid showed interactions with E protein at chain A: Arg 61, chain B: Phe 23, chain B: Tyr 57, and chain C: Val 25. A direct hit at the key residues forming helices and turns led to E protein structure disruption.

Moreover, on account of linkages between the chains, indirect effects were also noticed. For example, the ligand, glycyrrhizic acid interacted with chain B: Tyr 57, chain C: Leu 28, and chain C: Leu 31, which promoted disorganization among protein integrity. Further validation was taken up with  $\beta$ -boswellic acid, which revealed similar interaction with chain C of the protein. It pronounced indirect effect, showing interaction and the unfavorable binding that led to the protein's disruption.

Apart from these compounds, commercially available drugs such as azithromycin, remdesivir, dexamethasone, thymoquinone, and

hydroxychloroquine, which have been proposed as remedies against COVID-19, were also taken up as reference/control and compared with the twelve selected compounds screened in this work (Khan and Al-Balushi, 2021; Sharma et al., 2020). The present study showed that two medicinal compounds viz. glycyrrhizic acid and β-boswellic acid had greater efficacy and far better binding affinity than the five currently employed drugs for inhibiting COVID-19 (Yu et al., 2021). The binding affinity of presently used drugs in the treatment of COVID-19 are as follows; azithromycin (-5.9 kcal/mol), hydroxychloroquine (-6.4 kcal/mol), remdesivir (-7.7 kcal/mol), dexamethasone (-7.2 kcal/mol) and thymoquinone (-5.3 kcal/mol). Whereas our findings highlighted that out of the proposed medicinal metabolites, two were the most favourable bioactive compounds, i.e., β-boswellic and glycyrrhizic acids (Alam et al., 2020; Roy and Menon, 2021). These compounds showed a binding affinity of -9.1 kcal/mol, which performed far better in docking studies.

Moreover, apart from the structural insights, we delved into the pharmacological aspects of these compounds. Assessment of these compounds as drug formulation was done by DruLiTo 1.0. Various pharmacological aspects were studied for each compound, and their potential as a drug molecule was examined. The evaluation based on the parameters is not absolute, but it will be used to calculate ligand molecules for drug-like properties quickly. Parameters that defined the drug-likeliness are judged on the following criteria such as Lipinski's Rule (ClogP  $\leq$ 5), MDDR like rules (No. of rings  $\geq$ 3; No. of rigid bonds  $\geq$ 18; No. of rotatable bonds  $\geq$ 6), Veber Rule (Rotatable bond count  $\leq$ 10; PSA  $\leq$ 140), Ghose Filter (logP { $-0.4\sim5.6$ }; MR (Molar Refractivity 40~); molecular weight (160~480); Number of Atoms (20~70); Polar Surface Area < 140), BBR rule (H-bond {8-10}; Molecular weight (400-500); No. of acids), CMC-50 like rule (AlogP {1.3~4.1}; Molar refractivity (70-110); molecular weight (230-390); Number of Atoms (30-55)) and QED (Quantitative estimation of drug-likeness). Fig. 5 presents the drug-likeness of these compounds based on the parameters as mentioned earlier.

ADMET properties assessment: In-silico ADMET screening of the selected compounds

Further, toxicological assessment of these compounds and their ADME (Absorption, distribution, metabolism, and excretion) profile was

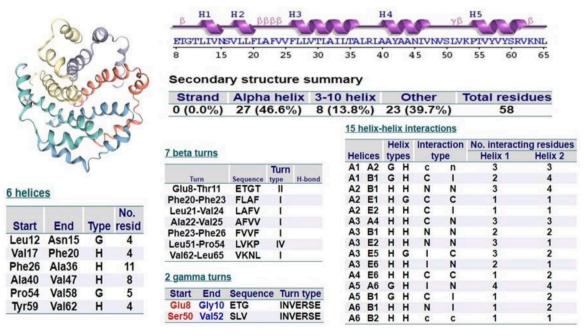


Fig. 2. Secondary structural assessment of target protein (SARS-CoV-2 E protein).

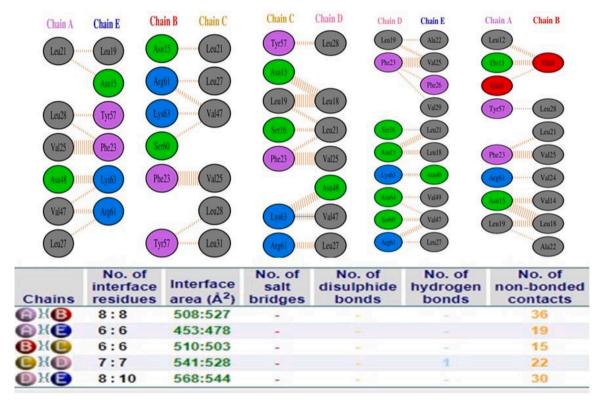


Fig. 3. Intramolecular interaction among pentameric chains of target protein (SARS-CoV-2 E protein).

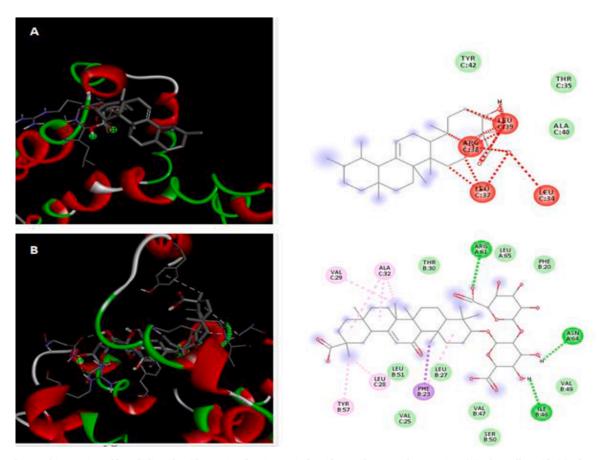


Fig. 4. Docking and interaction of best-fit ligands with proteins showing critical residues and nature of interactions (A) β-boswellic acid, (B) Glycyrrhizic acid.

inferred using vNN-ADMET prediction tool. This tool has a database of chemical compounds and their toxicological effects on the human body. With analysis, it gave a prediction of our target compounds. It examined their various toxicological properties like drug-induced liver injury (DILI), cytotoxicity, cardiotoxicity, mitochondrial toxicity (MMP), Blood-brain barrier (BBB), mutagenicity (AMES Test), Cytochrome P450 enzyme inhibition, and maximum recommended therapeutic dose (MRTD mg/day). Such evaluation explained the significance of tested compounds and their safe use for therapy. Fig. 6 showed a toxicological assessment of the screened compounds along with MRTD.

After validation, it was found that  $\beta$ -boswellic and glycyrrhizic acids were nontoxic. Considering the potency of  $\beta$ -boswellic and glycyrrhizic acids to mitigate efficiently in the present work, it seemed pertinent to study their *in-vitro* antiviral effects on SARS-CoV-2 E protein as well.

Effects of  $\beta$ -boswellic and glycyrrhizic on the structure of SARS-CoV-2 E protein

#### UV-visible spectral studies

Scans acquired from the range 200–700 nm led to prominent loss in the structural integrity of protein. The disruption was observed at 280 nm after treatment with both  $\beta\text{-}boswellic$  and glycyrrhizic acids in Fig 7.

#### Fluorescence spectroscopy

Fluorescence spectroscopy was applied to analyze drug–protein interactions by measuring the change of fluorescence emission intensity to decipher the quenching mechanism. The native  $\alpha\text{-helix}$  structure was seen to be destabilized in the presence of both  $\beta\text{-boswellic}$  and glycyrrhicic acids. The structural changes were notable on reaction with glycyrrhizic acid at 50  $\mu\text{g/ml}$ . Moreover, the E protein structure was completely disrupted by 100  $\mu\text{g/ml}$   $\beta\text{-boswellic}$  acid. The tertiary structural changes monitored by intrinsic fluorescence spectroscopy showed no emission spectra suggesting disruption in the microenvironment of the E protein.

As shown in Fig. 8, SARS-CoV-2 E showed a characteristic emission band at around 370 nm, whereas treated test samples i.e., treated with

 $\beta\text{-boswellic}$  and glycyrrhicic acids emitted almost no emission spectrum at the highest concentration. The fluorescence intensity of SARS-CoV-2 E was decreased progressively with the addition of increasing  $\beta\text{-boswellic}$  and glycyrrhicic acids concentrations (50 to 100 µg/ml), which indicated that these two medicinal metabolites interacted with SARS-CoV-2 E.

The presence of 100  $\mu g/ml$   $\beta$ -boswellic acid led to a maximum reduction in the fluorescence intensity. On the other hand, glycyrrhizic acid led to less changes in the tertiary structure of the SARS-CoV-2 E accompanied by blue shifts. Eventually with increasing concentration from 50 to 100  $\mu g/ml$  noticeable structural loss was observed by both these  $\beta$ -boswellic and glycyrrhicic acids. This might be the possible reason for the loss of activity in SARS-CoV-2 E. Effect on the active site of the SARS-CoV-2 E after binding by these acids was noticed (Gomaa et al., 2021; Sun et al., 2021; van de Sand et al., 2021). Hence the fluorescence quenching in both  $\beta$ -boswellic and glycyrrhicic acids indicated major changes in the immediate microenvironment of the SARS-CoV-2 E protein.

#### **Conclusions**

The present study was carried out to look for potential antiviral agents to combat COVID-19. Molecular docking studies were conducted to identify binding of medicinal metabolites with SARS-CoV-2 E protein. Out of screened compounds,  $\beta$ -boswellic acid (B. serrata) was found to be most suitable, along with glycyrrhizic acid (G. glabra). With the advantage of being a natural compound and certain pharmacological studies showed no harmful side effects, these novel compounds could be a good choice to be used for the treatment of COVID-19 patients. This study is in concordance to important antecedents with boswellic and glycyrrhinic acids against SARS-CoV2 and could be a probable treatment of COVID-19 (Caliebe et al., 2021; Gomaa et al., 2021; Li et al., 2021; Roy and Menon, 2021; Sun et al., 2021; van de Sand et al., 2021; Yu et al., 2021).

Hence, further investigations on their clinical use must be undertaken to validate  $\beta$ -boswellic and glycyrrhicic acids curative effects to

S. No	Compound	logp	Lipinskies Rule of Five	Ghose Filter	CMC- 50 like Rule	Vebers Rule	MDDR Like Rule	BBB Likeness Rule	Weighted QED	Unweighted QED
1	Aloesin	-1.116								
2	Azadiractin	-0.683								
3	Azithromycin	1.888								
4	Beta-boswellic Acid	10.438								
5	Cholecalciferol	9.896								
6	Cinnamaldehyde	1.96								
7	Dexamethasone	1.138								
8	Diosgenin	6.041								
9	Germacranolide	0.696								
10	Gingerols	2.437								
11	Glycyrrhizic acid	4.414								
12	Hydroxychloroquinone	1.548								
13	Isofraxidin	1.21								
14	Remdesivir	0.336								
15	Thymoquinone	1.102								
16	Tocopherol	10.514								

Fig. 5. Validation of screened compounds possessing drug-likeness properties.

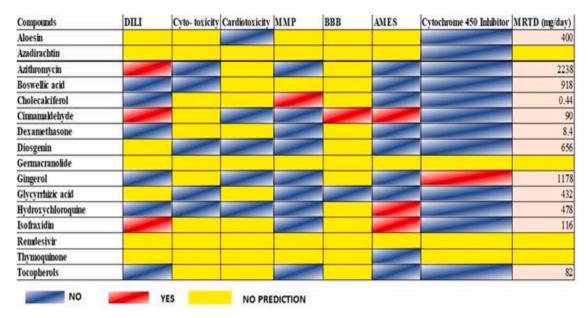


Fig. 6. Toxicological assessment of various parameters shown by tested compounds.

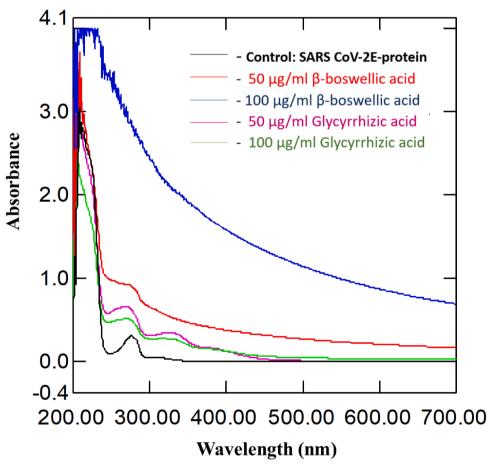


Fig. 7. Functionality loss and structural disruptions revealed by UV-vis spectra.

address the pandemic caused by SARS-CoV-2. A structural understanding of their effect on SARS-CoV-2 E, was achieved by UV and intrinsic fluorescence spectroscopy analyses. The UV and fluorescence spectra revealed that interaction with boswellic and glycyrrhicic acids caused microenvironmental and structural changes in E protein. In-vitro studies revealed the promising effect as potent SARS-CoV-2 E protein inhibitors.

These findings provide a rational basis for understanding drugs interaction with SARS-CoV-2 E for finding new compounds to cope with COVID-19 pandemic.

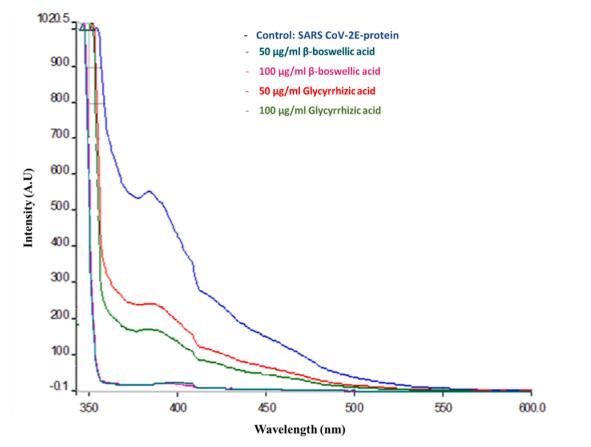


Fig. 8. Fluorescence spectra depicting the effect pronounced by various concentrations of Glycyrrhizic acid and  $\beta$ -boswellic acid on SARS-CoV-2 E protein.

## **Declaration of Competing interest**

The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

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#### Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.phyplu.2022.100241.

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